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(54) Title: HUMAN ANION TRANSPORTER GENES ATNOV

(57) Abstract

Methods for isolating ATnov genes are provided. The ATnov nucleic acid compositions find use in identifying homologous or related proteins and the DNA sequences encoding such proteins; in producing compositions that modulate the expression or function of the protein; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as identification of cell type based on expression, and the like.

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HUMAN ANION TRANSPORTER GENES

INTRODUCTION

Background

Endo- and xenobiotics are typically cleared from mammals via the liver, the primary site of drug metabolizing enzymes. Charged compounds, either endogenously or exogenously derived, are taken up by hepatocytes across the basolateral membrane, appropriately metabolized by the liver enzymes, trafficked through the cell, and then excreted across the canalicular membrane into the bile. These four steps are important in determining a patient's response to pharmaceutical agents.

Generation of bile flow is a regulated, ATP-dependent process and depends on the coordinated action of a number of transporter proteins in the sinusoidal and canalicular domains of the hepatocyte. Dysfunction of any of these proteins leads to retention of substrates, with conjugated hyperbilirubinemia or cholestasis as a result. In recent years many of the transport proteins involved in bile formation have been identified, cloned, and functionally characterized. The hepatocyte sinusoidal membrane contains transport proteins for the hepatic uptake of organic anions and cations and for the uptake of bile acids.

The Na+-independent organic anion transporter, OATP, resides on the basolateral surface of hepatocytes and mediates the uptake of a large number of amphipathic substrates, such as bromosulfophalein, bile acids, estrogen conjugates, neutral steroids, organic cations, cardiac glycosides, and peptidomimetic drugs. The human organic anion transporter, OATP, is expressed in multiple tissues, including brain, lung, liver, kidney, and testes, while a rat homolog of OATP is expressed only in liver and kidney (Bergwerk et al. (1996) Am. J. Physiol. 271: G231-G238). A prostaglandin transporter, hPGT, which shares significant homology with these organic anion transporters, is abundantly expressed (Lu et al. (1996) J. Clin. Invest. 98: 1142-1149).

Variations in transporter sequences may alter the kinetic properties of the protein. For example, inefficient clearance of substrates would result in an increased biological half-life, where drugs have an increased half-life and drug levels approach or reach toxic thresholds. Alternatively, over-efficient clearance of substrates could reduce the biological effectiveness of a drug. The identification of novel genes within these pathways provides additional targets for pharmacogenetic analysis, as well as a more thorough understanding of the biological process of drug clearance.

Relevant Literature

The molecular and functional characterization of an organic anion transporting polypeptide cloned from human liver, OATP, is described by Kullak-Ublick et al. (1995) Gastroenterology 109:1274-1282. Other cloned transporter genes are described by Noe et al. (1997) Proc. Natl. Acad. Sci. 94:10346-10350; and Jacquemin et al. (1994) Proc. Natl. Acad. Sci. 91:133-137.

The role of organic cation transporters in intestine, kidney, liver, and brain is reviewed by Koepsell (1998) Annu Rev Physiol 60:243-266. Canalicular multispecific organic anion transporter and the disposal of endo- and xenobiotics is reviewed by Elferink and Jansen (1994) Pharmac. Ther. 64:77-97.

Public EST sequences having sequence similarity with ATnov nucleic acids include: Genbank accessions nos. N49902 (ATnov2); N50005 (ATnov2); H62927 (ATnov3); H62893 (ATnov3); R29414 (ATnov3); AA382692 (ATnov3); T73863 (ATnov3); T74263 (ATnov3); T55488 (ATnov3).

SUMMARY OF THE INVENTION

Isolated nucleotide compositions and sequences are provided for ATnov genes. The ATnov nucleic acid compositions find use in identifying homologous or related genes; in producing compositions that modulate the expression or function of its encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as treatment of anion transporter defects, identification of cell type based on expression, and the like.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Nucleic acid compositions encoding ATnov anion transporters are provided. They are used in identifying homologous or related genes; in producing compositions that modulate the expression or function of the encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. The ATnov gene products are members of the anion transporter gene family, and have high degrees of homology at the amino acid level with known anion transporters.

CHARACTERIZATION OF ATNOV

The sequence data predict that the provided ATnov genes encode anion transporters. Characterization of organic ion transport across the cell membrane, in terms of substrates, binding and transport kinetics, is an important aspect of ATnov biology. A substrate, as used herein, is a chemical entity that is transported by an ATnov polypeptide, usually under normal physiological conditions. Substrates can be either endogenous substrates, i.e. substrates normally found within the natural environment, such as bile salts, or exogenous, i.e. substrates that are not normally found within the natural environment.

Substrate screening assays are used to determine the kinetics of a ATnov protein or peptide fragment on a substrate. Many suitable assays are known in the art, including the use of primary or cultured cells, genetically modified cells (e.g., where DNA encoding the ATnov polymorphism to be studied is introduced into the cell within an artificial construct), cell-free systems, e.g. recombinantly produced enzymes in a suitable buffer, or in animals, including human clinical trials (see, e.g. (1995) Burchell et al. Life Sci. 57:1819-1831, specifically incorporated herein by reference). Where genetically modified cells are used, since most cell lines do not express ATnov activity (liver cells lines being the exception), introduction of artificial construct for expression of the ATnov polymorphism into many human and non-human cell lines does not require additional modification of the host to inactivate endogenous ATnov expression/activity. Clinical trials may monitor serum, urine, etc. levels of the substrate or its metabolite(s).

Full length ion transporter cDNAs may be combined with proper vectors to form expression constructs of each individual transporter. Functional analyses of expressed transporters can be performed in heterologous systems, or by expression in mammalian cell lines. For expression analyses in heterologous systems such as Xenopus oocytes, synthetic mRNA is made through in vitro transcription of each transporter construct. mRNA is then injected into prepared oocytes and the cells allowed to express the transporter for several days. Candidate substrates may be labeled to provide a means of following movement across the membrane. Similarly, the requirements of a transporter for ATP, Na^+, etc. may be assessed. For an example of these techniques, see Kullak-Ublick et al. (1997) Gastroenterology 113(4):1295-1305.

Heterologous or mammalian cell lines expressing the novel transporters can be used to characterize small molecules and drugs that interact with the transporter. The same

experiments can be used to assay for novel compounds that interact with the expressed transporters.

ATNOV NUCLEIC ACID COMPOSITIONS

As used herein, the term "ATnov" is generically used to refer to any one of the provided nucleotide sequences as set forth in the SEQLIST. Of particular interest are the sequences, including polymorphisms, of ATnov3.1 and ATnov3.2. These sequences are provided as SEQ ID NO:3 (ATnov3.1), SEQ ID NO:5 (ATnov3.1), SEQ ID NO:7 (ATnov3.2) and SEQ ID NO:9 (ATnov3.2). The encoded polypeptides are provided as SEQ ID NO:4, 6, 8 and 10, respectively. The polymorphic variants are set forth in the sequences listings. These include a G or A polymorphism at nucleotide 487, resulting in an amino acid change of asp to asn. There is a polymorphism of C or T at nucleotide 670, which is silent with respect to the encoded polypeptide. A frameshift variant is found in the poly T stretch between positions 1705 and 1710, where the sequence contains either 5T or 6T. The 5T polymorphism results in a truncated polypeptide product of 542 amino acids (SEQ ID NO:4 and SEQ ID NO:8), while the 6T polymorphism encodes the full-length protein of 591 amino acids.

Also of interest are the genetic sequences of SEQ ID NO:1 (ATnov1) and SEQ ID NO:2 (ATnov2).

Where a specific ATnov sequence is intended, the numerical designation will be added. Nucleic acids encoding ATnov anion transporters may be cDNA or genomic DNA or a fragment thereof. The term "ATnov gene" shall be intended to mean the open reading frame encoding any of the provided ATnov polypeptides, introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

Novel nucleic acid compositions of the invention of particular interest comprise a sequence set forth in SEQ ID NO:1, 2, 3, 5, 7, 9 or an identifying sequence thereof. An "identifying sequence" is a contiguous sequence of residues at least about 10 nt to about 20 nt in length, usually at least about 50 nt to about 100 nt in length, that uniquely identifies a nucleic acid sequence, e.g., exhibits less than 90%, usually less than about 80% to about 85% sequence identity to any contiguous nucleotide sequence of more than about 20 nt.

Thus, the subject novel nucleic acid compositions include full length cDNAs or mRNAs that encompass an identifying sequence of contiguous nucleotides from SEQ ID NO:1, 2, 3, 5, 7, 9.

The nucleic acids of the invention also include nucleic acids having sequence similarity or sequence identity. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 10XSSC (0.9 M NaCl/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC. Sequence identity can be determined by hybridization under stringent conditions, for example, at 50°C or higher and 0.1XSSC (9 mM NaCl/0.9 mM sodium citrate). Hybridization methods and conditions are well known in the art, see U.S. Patent No. 5,707,829. Nucleic acids that are substantially identical to the provided nucleic acid sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided nucleic acid sequences (SEQ ID NO:1, 2, 3, 5, 7, 9) under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes can be any species.

Preferably, hybridization is performed using at least 15 contiguous nucleotides of SEQ ID NO:1, 2, 3, 5, 7, 9. The probe will preferentially hybridize with a nucleic acid or mRNA comprising the complementary sequence, allowing the identification and retrieval of the nucleic acids of the biological material that uniquely hybridize to the selected probe. Probes of more than 15 nucleotides can be used, e.g. probes of from about 18 nucleotides to not more than about 100 nucleotides, but 15 nucleotides generally represents sufficient sequence for unique identification.

The nucleic acids of the invention also include naturally occurring variants of the nucleotide sequences, e.g. degenerate variants, allelic variants, etc. Variants of the nucleic acids of the invention are identified by hybridization of putative variants with nucleotide sequences disclosed herein, preferably by hybridization under stringent conditions For example, by using appropriate wash conditions, variants of the nucleic acids of the invention can be identified where the allelic variant exhibits at most about 25-30% base pair mismatches relative to the selected nucleic acid probe. In general, allelic variants contain 5-25% base pair mismatches, and can contain as little as even 2-5%, or 1-2% base pair mismatches, as well as a single base-pair mismatch.

The invention also encompasses homologs corresponding to the nucleic acids of SEQ ID NO:1, 2, 3, 5, 7, 9, where the source of homologous genes can be any related

species within the same genus or group. Within a group, homologs have substantial sequence similarity, e.g. at least 75% sequence identity, usually at least 90%, more usually at least 95% between nucleotide sequences. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. A reference sequence will usually be at least about 18 contiguous nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al., J. Mol. Biol. (1990) 215:403-10.

In general, variants of the invention have a sequence identity greater than at least about 65%, preferably at least about 75%, more preferably at least about 85%, and can be greater than at least about 90% or more as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular). For the purposes of this invention, a preferred method of calculating percent identity is the Smith-Waterman algorithm, using the following. Global DNA sequence identity must be greater than 65% as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular) using an affine gap search with the following search parameters: gap open penalty, 12; and gap extension penalty, 1.

ATnov polymorphic sequences. It has been found that specific sites in the ATnov gene sequence are polymorphic, i.e. within a population, more than one nucleotide (G, A, T, C) is found at a specific position. Polymorphisms may provide functional differences in the genetic sequence, through changes in the encoded polypeptide, changes in mRNA stability, binding of transcriptional and translation factors to the DNA or RNA, and the like. The polymorphisms are also used as single nucleotide polymorphisms to detect association with, or genetic linkage to phenotypic variation in activity and expression of ATnov.

SNPs are generally biallelic systems, that is, there are two alleles that an individual may have for any particular marker. SNPs, found approximately every kilobase, offer the potential for generating very high density genetic maps, which will be extremely useful for developing haplotyping systems for genes or regions of interest, and because of the nature of SNPs, they may in fact be the polymorphisms associated with the disease phenotypes under study. The low mutation rate of SNPs also makes them excellent markers for studying complex genetic traits.

Single nucleotide polymorphisms are provided in the ATnov3 sequence listing The provided sequences also encompass the complementary sequence corresponding to any of the provided polymorphisms.

In order to provide an unambiguous identification of the specific site of a polymorphism, sequences flanking the polymorphic site are included in a probe for the region. It will be understood that there is no special significance to the length of non-polymorphic flanking sequence that is included, except to aid in positioning the polymorphism in the genomic sequence.

For screening purposes, hybridization probes of the polymorphic sequences may be used where both forms are present, either in separate reactions, spatially separated on a solid phase matrix, or labeled such that they can be distinguished from each other. Assays may utilize nucleic acids that hybridize to one or more of the described polymorphisms.

An array may include all or a subset of the ATnov3 polymorphisms. One or both polymorphic forms may be present in the array. Usually such an array will include at least 2 different polymorphic sequences, i.e. polymorphisms located at unique positions within the locus, and may include as many all of the provided polymorphisms. Arrays of interest may further comprise sequences, including polymorphisms, of other genetic sequences, particularly other sequences of interest for pharmacogenetic screening. The oligonucleotide sequence on the array will usually be at least about 12 nt in length, may be the length of the provided polymorphic sequences, or may extend into the flanking regions to generate fragments of 100 to 200 nt in length. For examples of arrays, see Ramsay (1998) Nat. Biotech. 16:40-44; Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

The subject nucleic acids can be cDNAs or genomic DNAs, as well as fragments thereof, particularly fragments that encode a biologically active gene product and/or are useful in the methods disclosed herein. The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when present, being removed by nuclear RNA splicing, to create a continuous open reading frame encoding a polypeptide of the invention.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the

introns that are normally present in a native chromosome. It can further include the 3' and 5' untranslated regions found in the mature mRNA. It can further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' and 3' end of the transcribed region. The genomic DNA can be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' and 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for expression.

The nucleic acid compositions of the subject invention can encode all or a part of the subject polypeptides. Double or single stranded fragments can be obtained from the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. Isolated nucleic acids and nucleic acid fragments of the invention comprise at least about 15 up to about 100 contiguous nucleotides, or up to the complete sequence provided in SEQ ID NO:1, 2, 3, 5, 7 or 9. For the most part, fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and up to at least about 50 contiguous nt in length or more.

Probes specific to the nucleic acids of the invention can be generated using the nucleic acid sequences disclosed in SEQ ID NO:1, 2, 3, 5, 7 or 9 and the fragments as described above. The probes can be synthesized chemically or can be generated from longer nucleic acids using restriction enzymes. The probes can be labeled, for example, with a radioactive, biotinylated, or fluorescent tag. Preferably, probes are designed based upon an identifying sequence of a nucleic acid of one of SEQ ID NO:1, 2, 3, 5, 7 or 9. More preferably, probes are designed based on a contiguous sequence of one of the subject nucleic acids that remain unmasked following application of a masking program for masking low complexity (e.g., XBLAST) to the sequence., i.e. one would select an unmasked region, as indicated by the nucleic acids outside the poly-n stretches of the masked sequence produced by the masking program.

The nucleic acids of the subject invention are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the nucleic acids, either as DNA or RNA, will be obtained substantially free of other naturally-occurring nucleic acid sequences, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", e.g., flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

The nucleic acids of the invention can be provided as a linear molecule or within a circular molecule. They can be provided within autonomously replicating molecules (vectors) or within molecules without replication sequences. They can be regulated by their own or by other regulatory sequences, as is known in the art. The nucleic acids of the invention can be introduced into suitable host cells using a variety of techniques which are available in the art, such as transferrin polycation-mediated DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated DNA transfer, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, gene gun, calcium phosphate-mediated transfection, and the like.

The subject nucleic acid compositions can be used to, for example, produce polypeptides, as probes for the detection of mRNA of the invention in biological samples (e.g., extracts of cells) to generate additional copies of the nucleic acids, to generate ribozymes or antisense oligonucleotides, and as single stranded DNA probes or as triple-strand forming oligonucleotides. The probes described herein can be used to, for example, determine the presence or absence of the nucleic acid sequences as shown in SEQ ID NO:1, 2, 3, 5, 7 or 9 or variants thereof in a sample.

The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where ATnov genes are expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms in the promoter regions are useful for determining natural variations in expression, particularly those that may be associated with disease.

Alternatively, mutations may be introduced into the promoter regions to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) Mol Med 1: 194-205; Mortlock et al. (1996) Genome Res. 6: 327-33; and Joulin and Richard-Foy (1995) Eur J Biochem 232: 620-626.

The regulatory sequences may be used to identify cis acting sequences required for transcriptional or translational regulation of ATnov expression, especially in different tissues or stages of development, and to identify cis acting sequences and trans acting factors that regulate or mediate ATnov expression. Such transcription or translational control regions

may be operably linked to a ATnov gene in order to promote expression of wild type or altered ATnov or other proteins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

Double or single stranded fragments may be obtained of the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. For the most part, DNA fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and may be at least about 50 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 100 nt are useful for production of the encoded polypeptide. For use in amplification reactions, such as PCR, a pair of primers will be used. The exact composition of the primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for the selection of primer sequences are generally known, and are available in commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

The DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide sequences, as genomic DNA or RNA, is well established in the literature and does not require elaboration here. DNA or mRNA is isolated from a cell sample. The mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, the mRNA sample is separated by gel electrophoresis, transferred to a suitable support, e.g. nitrocellulose, nylon, etc., and then probed with a fragment of the subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, in situ hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to the subject sequence is indicative of ATnov gene expression in the sample.

The sequence of an ATnov gene, including flanking promoter regions and coding regions, may be mutated in various ways known in the art to generate targeted changes in promoter strength, sequence of the encoded protein, etc. The DNA sequence or protein product of such a mutation will usually be substantially similar to the sequences provided

herein, i.e. will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions. Deletions may further include larger changes, such as deletions of a domain or exon. Other modifications of interest include epitope tagging, e.g. with the FLAG system, HA, etc. For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used.

Techniques for in vitro mutagenesis of cloned genes are known. Examples of protocols for site specific mutagenesis may be found in Gustin et al., Biotechniques 14:22 (1993); Barany, Gene 37:111-23 (1985); Colicelli et al., Mol Gen Genet 199:537-9 (1985); and Prentki et al., Gene 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook et al., Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp. 15.3-15.108; Weiner et al., Gene 126:35-41 (1993); Sayers et al., Biotechniques 13:592-6 (1992); Jones and Winistorfer, Biotechniques 12:528-30 (1992); Barton et al., Nucleic Acids Res 18:7349-55 (1990); Marotti and Tomich, Gene Anal Tech 6:67-70 (1989); and Zhu, Anal Biochem 177:120-4 (1989). Such mutated genes may be used to study structure-function relationships of ATnov polypeptides, or to alter properties of the protein that affect its function or regulation.

Genetic polymorphisms, either naturally occurring or introduced as described above, are useful in screening for altered transport or metabolism of ATnov substrates. For example, variant alleles may affect the pharmacokinetic parameters of substrates. A drug□s volume of distribution, clearance, and the derived parameter, half-life, are particularly important, as they determine the degree of fluctuation between a maximum and minimum plasma concentration during a dosage interval, the magnitude of steady state concentration and the time to reach steady state plasma concentration upon chronic dosing. Parameters derived from in vivo drug administration are useful in determining the clinical effect of a particular ATnov genotype.

ATNOV POLYPEPTIDES

The subject gene may be employed for producing all or portions of ATnov polypeptides. Fragments of interest include the glycosylation sites, transmembrane domains, ATP binding regions, the substrate binding sites, etc. Such domains will usually include at least about 20 amino acids of the provided sequence, more usually at least about 50 amino acids, and may include 100 amino acids or more, up to the complete domain.

Binding contacts may be comprised of non-contiguous sequences, which are brought into proximity by the tertiary structure of the protein. The sequence of such fragments may be modified through manipulation of the coding sequence, as described above. Truncations may be performed at the carboxy or amino terminus of the fragment, e.g. to determine the minimum sequence required for biological activity.

A subset of the provided nucleic acid polymorphisms in ATnov3 confer a change in the corresponding amino acid sequence, as previously described. Using the amino acid sequence provided in SEQ ID NO:3 as a reference, the amino acid polymorphisms of the invention include asn□asp, pos. 130; and a frameshift at position 537 resulting in a truncated protein of 542 amino acids. Polypeptides comprising at least one of the provided polymorphisms (ATnov3^{v} polypeptides) are of interest. The term "ATnov3^{v} polypeptides" as used herein includes complete ATnov protein forms, e.g. such splicing variants as known in the art, and fragments thereof, which fragments may comprise short polypeptides, epitopes, functional domains; binding sites; etc.; and including fusions of the subject polypeptides to other proteins or parts thereof. Polypeptides will usually be at least about 8 amino acids in length, more usually at least about 12 amino acids in length, and may be 20 amino acids or longer, up to substantially the complete protein.

For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or constitutive, where the coding region is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to an ATnov gene, or may be derived from exogenous sources.

The peptide may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as E. coli, B. subtilis, S. cerevisiae, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, e.g. COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the ATnov gene in eukaryotic cells, where the ATnov protein will benefit from native folding and post-translational modifications. Small peptides can also be synthesized in the laboratory. Peptides that are subsets of the complete ATnov sequence may be used to identify and investigate parts of the protein important for function, or to raise antibodies directed against these regions.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique. The purified protein will generally be at least about 80% pure, preferably at least about 90% pure, and may be up to and including 100% pure. Pure is intended to mean free of other proteins, as well as cellular debris.

The expressed ATnov polypeptides are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Antibodies may be raised to the wild-type or variant forms of ATnov. Antibodies may be raised to isolated peptides corresponding to these domains, or to the native protein.

Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see Monoclonal Antibodies: A Laboratory Manual, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in E. coli, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to in vivo immunization as a method of raising antibodies include binding to phage "display" libraries, usually in conjunction with in vitro affinity maturation.

ATNOV GENOTYPING

The subject nucleic acid and/or polypeptide compositions may be used in genotyping and to screen for the presence of polymorphisms in the sequence, or variation in the expression of the subject genes. Genotyping may be performed to determine whether a particular polymorphisms is associated with a disease state or genetic predisposition to a disease state, particularly diseases associated with liver disorders.

Genotyping may also be performed for pharmacogenetic analysis to assess the association between an individual's genotype and that individual's ability to react to a therapeutic agent. Differences in substrate transport to relevant cells can lead to toxicity or therapeutic failure. Relationships between polymorphisms in transporter expression or specificity can be used to optimize therapeutic dose administration.

ATnov genotyping is performed by DNA or RNA sequence and/or hybridization analysis of any convenient sample from a patient, e.g. biopsy material, blood sample, scrapings from cheek, etc. A nucleic acid sample from an individual is analyzed for the presence of polymorphisms in ATnov, particularly those that affect the activity, responsiveness or expression of ATnov. Specific sequences of interest include any polymorphism that leads to changes in basal expression in one or more tissues, to changes in the modulation of ATnov expression, or alterations in ATnov specificity and/or activity.

The effect of a polymorphism in ATnov gene sequence on the response to a particular agent may be determined by in vitro or in vivo assays. Such assays may include monitoring during clinical trials, testing on genetically defined cell lines, etc. The response of an individual to the agent can then be predicted by determining the ATnov genotype with respect to the polymorphism. Where there is a differential distribution of a polymorphism by racial background, guidelines for drug administration can be generally tailored to a particular ethnic group.

Biochemical studies may be performed to determine whether a sequence polymorphism in a ATnov coding region or control regions is associated with disease. Disease associated polymorphisms may include deletion or truncation of the gene, mutations that alter expression level, that affect the specificity or transport kinetics of the transporter, etc.

A number of methods are available for analyzing nucleic acids for the presence of a specific sequence. Where large amounts of DNA are available, genomic DNA is used directly. Alternatively, the region of interest is cloned into a suitable vector and grown in sufficient quantity for analysis. The nucleic acid may be amplified by conventional techniques, such as the polymerase chain reaction (PCR), to provide sufficient amounts for analysis. The use of the polymerase chain reaction is described in Saiki et al. (1985) Science 239:487, and a review of current techniques may be found in Sambrook et al. Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2□14.33. Amplification may be used to determine whether a polymorphism is present, by using a primer that is

specific for the polymorphism. Alternatively, various methods are known in the art that utilize oligonucleotide ligation as a means of detecting polymorphisms, for examples see Delahunty et al. (1996) Am. J. Hum. Genet.58:1239-1246.

A detectable label may be included in an amplification reaction. Suitable labels include fluorochromes, e.g. fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'-dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX), 6-carboxy-2',4',7',4,7-hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6-carboxyrhodamine (TAMRA), radioactive labels, e.g. ^{32}P, ^{35}S, ^{3}H; etc. The label may be a two stage system, where the amplified DNA is conjugated to biotin, haptens, etc. having a high affinity binding partner, e.g. avidin, specific antibodies, etc., where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

The sample nucleic acid, e.g. amplified or cloned fragment, is analyzed by one of a number of methods known in the art. The nucleic acid may be sequenced by dideoxy or other methods. Hybridization with the variant sequence may also be used to determine its presence, by Southern blots, dot blots, etc. The hybridization pattern of a control and variant sequence to an array of oligonucleotide probes immobilised on a solid support, as described in U.S. 5,445,934, or in WO95/35505, may also be used as a means of detecting the presence of variant sequences. Single strand conformational polymorphism (SSCP) analysis, denaturing gradient gel electrophoresis (DGGE), mismatch cleavage detection, and heteroduplex analysis in gel matrices are used to detect conformational changes created by DNA sequence variation as alterations in electrophoretic mobility. Alternatively, where a polymorphism creates or destroys a recognition site for a restriction endonuclease (restriction fragment length polymorphism, RFLP), the sample is digested with that endonuclease, and the products size fractionated to determine whether the fragment was digested. Fractionation is performed by gel or capillary electrophoresis, particularly acrylamide or agarose gels.

In one embodiment of the invention, an array of oligonucleotides are provided, where discrete positions on the array are complementary to one or more of the provided polymorphic sequences, e.g. oligonucleotides of at least 12 nt, frequently 20 nt, or larger, and including the sequence flanking the polymorphic position. Such an array may comprise

a series of oligonucleotides, each of which can specifically hybridize to a different polymorphism. For examples of arrays, see Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

Screening for polymorphisms in ATnov may be based on the functional or antigenic characteristics of the protein. Protein truncation assays are useful in detecting deletions that may affect the biological activity of the protein. Various immunoassays designed to detect polymorphisms in ATnov proteins may be used in screening. Where many diverse genetic mutations lead to a particular disease phenotype, functional protein assays have proven to be effective screening tools. The activity of the encoded ATnov protein as a anion transporter may be determined by comparison with the wild-type protein.

Antibodies specific for a ATnov may be used in staining or in immunoassays. Samples, as used herein, include biological fluids such as semen, blood, cerebrospinal fluid, tears, saliva, lymph, dialysis fluid and the like; organ or tissue culture derived fluids; and fluids extracted from physiological tissues. Also included in the term are derivatives and fractions of such fluids. The cells may be dissociated, in the case of solid tissues, or tissue sections may be analyzed. Alternatively a lysate of the cells may be prepared.

Diagnosis may be performed by a number of methods to determine the absence or presence or altered amounts of normal or abnormal ATnov polypeptides in patient cells. For example, detection may utilize staining of cells or histological sections, performed in accordance with conventional methods. The antibodies of interest are added to the cell sample, and incubated for a period of time sufficient to allow binding to the epitope, usually at least about 10 minutes. The antibody may be labeled with radioisotopes, enzymes, fluorescers, chemiluminescers, or other labels for direct detection. Alternatively, a second stage antibody or reagent is used to amplify the signal. Such reagents are well known in the art. For example, the primary antibody may be conjugated to biotin, with horseradish peroxidase-conjugated avidin added as a second stage reagent. Alternatively, the secondary antibody conjugated to a flourescent compound, e.g. flourescein, rhodamine, Texas red, etc. Final detection uses a substrate that undergoes a color change in the presence of the peroxidase. The absence or presence of antibody binding may be determined by various methods, including flow cytometry of dissociated cells, microscopy, radiography, scintillation counting, etc.

MODULATION OF GENE EXPRESSION

The ATnov genes, gene fragments, or the encoded protein or protein fragments are useful in gene therapy to treat disorders associated with ATnov defects. Expression vectors may be used to introduce the ATnov gene into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about one day, more usually for a period of at least about several days to several weeks.

The gene or ATnov protein may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of vesicles. Jet injection may also be used for intramuscular administration, as described by Furth et al. (1992) Anal Biochem 205:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang et al. (1992) Nature 356:152-154), where gold microprojectiles are coated with the ATnov or DNA, then bombarded into skin cells.

Antisense molecules can be used to down-regulate expression of ATnov in cells. The anti-sense reagent may be antisense oligonucleotides (ODN), particularly synthetic ODN having chemical modifications from native nucleic acids, or nucleic acid constructs that express such anti-sense molecules as RNA. The antisense sequence is complementary to the mRNA of the targeted gene, and inhibits expression of the targeted gene products. Antisense molecules inhibit gene expression through various mechanisms, e.g. by reducing the amount of mRNA available for translation, through activation of RNAse H, or steric hindrance. One or a combination of antisense molecules may be administered, where a combination may comprise multiple different sequences.

Antisense molecules may be produced by expression of all or a part of the target gene sequence in an appropriate vector, where the transcriptional initiation is oriented such that an antisense strand is produced as an RNA molecule. Alternatively, the antisense molecule is a synthetic oligonucleotide. Antisense oligonucleotides will generally be at least about 7, usually at least about 12, more usually at least about 20 nucleotides in length, and not more than about 500, usually not more than about 50, more usually not more than about

35 nucleotides in length, where the length is governed by efficiency of inhibition, specificity, including absence of cross-reactivity, and the like. It has been found that short oligonucleotides, of from 7 to 8 bases in length, can be strong and selective inhibitors of gene expression (see Wagner et al. (1996) Nature Biotechnology 14:840-844).

A specific region or regions of the endogenous sense strand mRNA sequence is chosen to be complemented by the antisense sequence. Selection of a specific sequence for the oligonucleotide may use an empirical method, where several candidate sequences are assayed for inhibition of expression of the target gene in an in vitro or animal model. A combination of sequences may also be used, where several regions of the mRNA sequence are selected for antisense complementation.

Antisense oligonucleotides may be chemically synthesized by methods known in the art (see Wagner et al. (1993) supra. and Milligan et al., supra.) Preferred oligonucleotides are chemically modified from the native phosphodiester structure, in order to increase their intracellular stability and binding affinity. A number of such modifications have been described in the literature, which alter the chemistry of the backbone, sugars or heterocyclic bases.

Among useful changes in the backbone chemistry are phosphorothioates; phosphorodithioates, where both of the non-bridging oxygens are substituted with sulfur; phosphoroamidites; alkyl phosphotriesters and boranophosphates. Achiral phosphate derivatives include 3□-O□-5□-S-phosphorothioate, 3□-S-5□-O-phosphorothioate, 3□-CH2-5□-O-phosphonate and 3□-NH-5□-O-phosphoroamidate. Peptide nucleic acids replace the entire ribose phosphodiester backbone with a peptide linkage. Sugar modifications are also used to enhance stability and affinity. The □-anomer of deoxyribose may be used, where the base is inverted with respect to the natural □-anomer. The 2□-OH of the ribose sugar may be altered to form 2□-O-methyl or 2□-O-allyl sugars, which provides resistance to degradation without comprising affinity. Modification of the heterocyclic bases must maintain proper base pairing. Some useful substitutions include deoxyuridine for deoxythymidine; 5-methyl-2□-deoxycytidine and 5-bromo-2□-deoxycytidine for deoxycytidine. 5- propynyl-2□-deoxyuridine and 5-propynyl-2□-deoxycytidine have been shown to increase affinity and biological activity when substituted for deoxythymidine and deoxycytidine, respectively.

As an alternative to anti-sense inhibitors, catalytic nucleic acid compounds, e.g. ribozymes, anti-sense conjugates, etc. may be used to inhibit gene expression. Ribozymes may be synthesized in vitro and administered to the patient, or may be encoded on an

expression vector, from which the ribozyme is synthesized in the targeted cell (for example, see International patent application WO 9523225, and Beigelman et al. (1995) Nucl. Acids Res 23:4434-42). Examples of oligonucleotides with catalytic activity are described in WO 9506764. Conjugates of anti-sense ODN with a metal complex, e.g. terpyridylCu(II), capable of mediating mRNA hydrolysis are described in Bashkin et al. (1995) Appl Biochem Biotechnol 54:43-56.

GENETICALLY ALTERED CELL OR ANIMAL MODELS FOR ATNOV FUNCTION

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. Transgenic animals may be made through homologous recombination, where the normal ATnov locus is altered. Alternatively, a nucleic acid construct is randomly integrated into the genome. Vectors for stable integration include plasmids, retroviruses and other animal viruses, YACs, and the like.

The modified cells or animals are useful in the study of ATnov function and regulation. For example, a series of small deletions and/or substitutions may be made in the ATnov gene to determine the role of different transmembrane domains, of ATP catalysis, etc. Of interest are the use of ATnov to construct transgenic animal models where expression of ATnov is specifically reduced or absent. Specific constructs of interest include anti-sense ATnov, which will block ATnov expression, expression of dominant negative ATnov mutations, etc. One may also provide for expression of the ATnov gene or variants thereof in cells or tissues where it is not normally expressed or at abnormal times of development.

DNA constructs for homologous recombination will comprise at least a portion of the ATnov gene with the desired genetic modification, and will include regions of homology to the target locus. DNA constructs for random integration need not include regions of homology to mediate recombination. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are known in the art. For various techniques for transfecting mammalian cells, see Keown et al. (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g. mouse, rat, guinea pig, etc. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES or embryonic cells have been transformed, they may be used to

produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the construct. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting offspring screened for the construct. By providing for a different phenotype of the blastocyst and the genetically modified cells, chimeric progeny can be readily detected.

The chimeric animals are screened for the presence of the modified gene and males and females having the modification are mated to produce homozygous progeny. If the gene alterations cause lethality at some point in development, tissues or organs can be maintained as allogeneic or congenic grafts or transplants, or in in vitro culture. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, etc. The transgenic animals may be used in functional studies, drug screening, etc.

TESTING OF ATNOV FUNCTION and RESPONSES

Anion transporters such as ATnov polypeptides are involved in multiple biologically important processes. Pharmacological agents designed to affect only specific transporter subtypes are of particular interest. The subject polypeptides may be used to test the specificity of novel compounds, and of analogs and derivatives of compounds known to be substrates, or to act on anion transporters.

Drug screening may be performed using an in vitro model, a genetically altered cell or animal, or purified ATnov protein. One can identify ligands or substrates that bind to, modulate or mimic the action of ATnov. Drug screening identifies agents that provide a replacement for ATnov function in abnormal cells. Of particular interest are screening assays for agents that have a low toxicity for human cells. A wide variety of assays may be used for this purpose, including monitoring cellular excitation and conductance, labeled in vitro protein-protein binding assays, electrophoretic mobility shift assays, immunoassays for protein binding, and the like. The purified protein may also be used for determination of

three-dimensional crystal structure, which can be used for modeling intermolecular interactions.

The term "agent" as used herein describes any molecule, e.g. protein or pharmaceutical, with the capability of altering or mimicking the physiological function of ATnov polypeptide. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, though typically they are organic molecules, preferably small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical means, and may be used to produce combinatorial libraries. Known pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, amidification, etc. to produce structural analogs.

Where the screening assay is a binding assay, one or more of the molecules may be joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific

binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc. that are used to facilitate optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40°C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

The compounds having the desired pharmacological activity may be administered in a physiologically acceptable carrier to a host in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, etc. Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of therapeutically active compound in the formulation may vary from about 0.1-100 wt.%. The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the like. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for oral and topical use can be used to make up compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents.

EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (e.g. amounts, temperature, concentrations, etc.) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

Example 1

Three novel members of the OATP gene family, which are expressed in liver tissue, were cloned. These genes were isolated using trapped exons obtained from large-scale exon trapping of chromosome 12. The three anion transporters reported here are 70-80% identical to each other over the predicted protein sequence, and are each 40% identical to the reported OATP protein sequence (Kullack-Ublick et al., (1995) Gastroenterology 109:1274-1282). The chromosomal location of these three anion transporters, along with the mapping of OATP, suggests this gene-family is clustered on 12p12.

Materials and Methods

cDNA Isolation. cDNA clones were isolated using the GeneTrapper system (Gibco-BRL). PCR primers within the trapped exons were used to detect which plasmid cDNA libraries contained the gene of interest. Oligonucleotide probes were designed: (SEQ ID NO:7) C12B_120: GGGGCTCTGATTGATACAACGTG ; (SEQ ID NO:8) C12C_151: ACTGTGGCACACGTGGGTCATGTAGGACAT) and the process proceeded according to the supplied protocol. cDNA clones were sequenced on an ABI 377 according to standard methods. The Primer Island Transposition kit was used according to the supplied protocol. Sequences were analyzed, edited, and assembled using the Sequencher software (Gene Codes).

Radiation Hybrid Mapping. RH mapping was achieved using the Stanford G3 panel DNAs (Research Genetics). DNA was aliquoted into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. 20 µl PCR reactions with standard conditions, 2.5

mM $MgCl_2$, Taq Gold, and an annealing temperature of 60°C (for ATnov1 and 2) or 55°C (for ATnov3) were used to detect expression. The assays were done in duplicate and results were scored and map positions determined via the RH server at Stanford University <http://www-shgc.stanford.edu/RH/G3index.html>.

RT-PCR. RT-PCR was utilized to characterize the expression pattern of the novel anion transporters. This approach used RNA from 30 different tissues to generate first strand cDNA. Total RNA was purchased (Clontech, Invitrogen) and used to synthesize first strand cDNA using M-MLV reverse transcriptase and the supplied buffer (Gibco-BRL). The 20 µl reaction contained 5 µg total RNA, 100 ng of random primers, 10 mM DTT, 0.5 mM each dNTP, and an RNAse inhibitor (Gibco-BRL). Identical reactions were set up without reverse transcriptase to control for DNA contamination in the RNA samples. The synthesis reaction proceeded for 1 hour at 37°C followed by 10 minutes at 95°C. These cDNAs, along with control cDNA synthesis reactions without reverse transcriptase, were diluted 1:5 and 2 µl of each sample were arrayed into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. The cDNAs were tested with primers with defined expression patterns to verify the presence of amplifiable cDNA from each tissue. Gene-specific primers were used to amplify the cDNAs in 20 µl PCR reactions with standard conditions, 2.5 mM $MgCl_2$, Taq Gold, and an appropriate annealing temperature.

This approach provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner and provides qualitative analysis of gene expression only. Modifications can be employed, such as the use of internal control primers, limited cycling parameters, and dilution series to convert this to a quantitative experiment.

Primers for ATnov1

RH primers (SEQ ID NO:11) CTGCTGCCAACTAACATTGC
(SEQ ID NO:12) CACACACTAACCATGCCTCT
237 bp product

RT-PCR primers (SEQ ID NO:13) TCCAGTCATTGGCTTTGCAC
(SEQ ID NO:14) AAGAACCAATAAAGCTGCTTACT
413 bp product

Primers for ATnov2

RH primers (SEQ ID NO:15) GTGTTTGCTAGCCACCTTGA

(SEQ ID NO:16) GGCAACACTTCCTCAAAGTG

196 bp product

RT-PCR primers (SEQ ID NO:17) GATGCTTTCCTCTGTGCAGT

(SEQ ID NO:18) CCTTCAAGCCGAAGAAGGCT

259 bp product

Primers for ATnov3

RH primers (SEQ ID NO:19) AGGAGTTCCTGGTCCTTTCA

(SEQ ID NO:20) CAAGCTAGACTTCAGGCCTT

137 bp product

RT-PCR primers (SEQ ID NO:21) GAGGAATTCTAGCTCCAATATATT

(SEQ ID NO:22) GTCCTACATGACCCACGTGTG

96 bp product

Results

cDNA Isolation. Large-scale exon trapping was completed across a chromosome 12 cosmid library. Approximately 2400 exons were sequenced and analyzed by BLAST algorithms to identify exons with potentially interesting homologies. Two different exons, C12B_120 and C12C_151 were identified that were 87% identical to each other and ~68% identical to a cloned organic anion transporter, OATP (Kullak-Ubrick et al. (1995) Gastroenterology 109: 1274-1282.), at the DNA level and ~78% similar at the amino acid level. Full-length cDNA clones were isolated using GeneTrapper (Gibco-BRL) from a liver cDNA library (Gibco-BRL). The resulting clones, the largest being up to 3.0 kb, were end-sequenced using vector primers. If the end sequences provided insufficient coverage of the cDNA clones, a transposon approach was used to complete the sequence of the cDNA clone.

The cDNA clones isolated with C12B_120 yielded two different sequence contigs, ATnov1 and ATnov2, which were ~89% identical to each other. ATnov2 is identical to C12B_120. cDNA clones isolated with C12C_151 generated a sequence contig, ATnov3, that was ~86% identical to the first two contigs. Conceptual translations yielded predicted proteins of 688-704 amino acids in length. A multiple alignment of these three proteins is shown in figure 1. These genes also show significant homology to a human organic anion

transporter OATP (~40% identity, 60% similarity) and to a human prostaglandin transporter (~32% identity, 51% similarity) over the length of the predicted proteins.

Chromosomal Localization. The exon trapped products used in the cDNA screens were trapped from a chromosome 12 cosmid library, suggesting that at least ATnov2 and ATnov3 map to chromosome 12. OATP had been previously reported to map to chromosome 12 (Kullak-Ubrick et al., supra.) Radiation hybrid mapping was used to confirm the localization of these to chromosome 12, as well as to map them and ATnov1 to a specific region on the chromosome. The Stanford G3 panel showed linkage of all four of these genes to the marker GATA91H01, which is extrapolated to a cytogenetic location of 12p12.

Expression Analysis. OATP is expressed in multiple tissues, including brain, lung, liver, kidney, and testes (Kullak-Ubrick et al., supra.) RT-PCR was utilized to characterize the expression pattern of the novel anion transporters. This approach used RNA from 30 different tissues to generate first strand cDNA. These cDNAs were arrayed, along with control cDNA synthesis reactions without reverse transcriptase, into 96-well trays, dried and stored until needed. This resource provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner. RT-PCR in this fashion allows for qualitative analysis of gene expression only.

PCR was performed on these plates with gene-specific primers for each of the ATnov genes. ATnov1 is expressed in fetal and adult liver; ATnov2 is expressed in adult liver and mammary gland; ATnov3 is expressed in fetal liver, adult liver, brain, adipose tissue, skin, and testes.

The predicted positions of transmembrane domains in the ATnov3 polypeptide are as follows:

	ATnov3
Transmembrane domain 1	29-45
Transmembrane domain 2	94-104
Transmembrane domain 3	169-189
Transmembrane domain 4	207-227
Transmembrane domain 5	259-279
Transmembrane domain 6	336-356

Transmembrane domain 7	376-396
Transmembrane domain 8	410-430
Transmembrane domain 9	481-501
Transmembrane domain 10	537-557
Transmembrane domain 11	581-601
Transmembrane domain 12	627-647

These novel members of the organic anion transporter family are expressed in the liver. Based on homology to another organic anion transporter, they are likely to be present on the basolateral surface of the hepatocytes and mediate the uptake of both xenobiotics and endogenous compounds for metabolism by the cytochrome p450s, glucuronosyl transferases, and other metabolic enzymes known to be present in the liver. The ATnov genes are all expressed in the liver, with ATnov 2 and 3 also being expressed in a limited number of other tissues. The RT-PCR approach described herein has a high level of sensitivity, with the ability to detect a control transcript diluted down to an expression level equivalent to a frequency of $1/10^7$.

The map positions of these anion transporters suggest that they lie adjacent to each other on the proximal short arm of chromosome 12. The anion transporters described herein are only ~89% identical to each other at the DNA level, suggesting that these genes arose via a recombination mechanism, but have since diverged sufficiently such that it is unlikely that these genes are polymorphic within a given population.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid encoding a mammalian ATnov protein.

2. An isolated nucleic acid according to Claim 1, wherein said ATnov protein has the amino acid sequence of SEQ ID NO:4, 6, 8, or 10.

3. An isolated nucleic acid according to Claim 1, wherein said ATnov protein has an amino acid sequence that is substantially identical to the amino acid sequence of SEQ ID NO:4, 6, 8, or 10.

4. An isolated nucleic acid according to Claim 1, comprising the nucleotide sequence as set forth in SEQ ID NO:1,2, 3, 5, 7, or 9.

5. An isolated nucleic acid that hybridizes under stringent conditions to the nucleic acid sequence of claim 4.

6. An expression cassette comprising a transcriptional initiation region functional in an expression host, a nucleic acid having a sequence of the isolated nucleic acid according to Claim 1 under the transcriptional regulation of said transcriptional initiation region, and a transcriptional termination region functional in said expression host.

7. A cell comprising an expression cassette according to Claim 6 as part of an extrachromosomal element or integrated into the genome of a host cell as a result of introduction of said expression cassette into said host cell, and the cellular progeny of said host cell.

8. A method for producing mammalian ATnov protein, said method comprising:

growing a cell according to Claim 7, whereby said mammalian ATnov protein is expressed; and

isolating said ATnov protein free of other proteins.

9. A purified polypeptide composition comprising at least 50 weight % of the protein present as a ATnov protein or a fragment thereof.

10. A monoclonal antibody binding specifically to an ATnov protein.

11. A non-human transgenic animal model for ATnov gene function wherein said transgenic animal comprises an introduced alteration in an ATnov gene.

12. The animal model of claim 11, wherein said animal is heterozygous for said introduced alteration.

13. The animal model of claim 12, wherein said animal is homozygous for said introduced alteration.

14. The animal model of claim 12, wherein said introduced alteration is a knockout of endogenous ATnov gene expression.

15. An isolated nucleic acid probe comprising an ATnov 3 sequence polymorphism, as part of other than a naturally occurring chromosome.

16. A nucleic acid probe according to Claim 15, wherein said probe is conjugated to a detectable marker.

17. An array of oligonucleotides comprising:
two or more probes for detection of ATnov3 locus polymorphisms.

SEQUENCE LISTING

<110> Miller, Andrew
Buckler, Alan
Laubert, Boris

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His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser Glu Asn Lys Lys Thr
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Cys Asp Glu Ser Gln Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr
465                 470                 475                 480
Tyr Ile Ser Pro Cys Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys
                485                 490                 495
Lys Pro Ile Val Phe Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu
            500                 505                 510
Gln Asn Arg Asn Tyr Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp
        515                 520                 525
Ala Cys Thr Arg Lys Phe Tyr Phe Leu Leu Gln Tyr Lys Ser
    530                 535                 540
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<210> 5
<211> 2800
<212> DNA
<213> H. sapiens

<220>
<221> CDS
<222> (100)...(2175)
<223> Coding sequence of ATnov3.1

<221> variation
<222> (1705)...(1710)
<223> Polymorphism of 5 or 6 T residues.

<221> variation
<222> (487)...(487)
<223> Polymorphism of A or G

<221> variation
<222> (670)...(670)
<223> Polymorphism of C or T

<400> 5

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gtggacttgt tgcagttgct gtaggattct aaatccaggt gattgtttca aactgagcat      60
caacaacaaa aacatttgta tgatatctat atttcaatc atg gac caa aat caa         114
                                          Met Asp Gln Asn Gln
                                           1               5

cat ttg aat aaa aca gca gag gca caa cct tca gag aat aag aaa aca         162
His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser Glu Asn Lys Lys Thr
                10                  15                  20
```

```
aga tac tgc aat gga ttg aag atg ttc ttg gca gct ctg tca ctc agc      210
Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala Ala Leu Ser Leu Ser
            25                  30                  35

ttt att gct aag aca cta ggt gca att att atg aaa agt tcc atc att      258
Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met Lys Ser Ser Ile Ile
        40                  45                  50

cat ata gaa cgg aga ttt gag ata tcc tct tct ctt gtt ggt ttt att      306
His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser Leu Val Gly Phe Ile
    55                  60                  65

gac gga agc ttt gaa att gga aat ttg ctt gtg att gta ttt gtg agt      354
Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val Ile Val Phe Val Ser
70                  75                  80                  85

tac ttt gga tcc aaa cta cat aga cca aag tta att gga atc ggt tgt      402
Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu Ile Gly Ile Gly Cys
                90                  95                  100

ttc att atg gga att gga ggt gtt ttg act gct ttg cca cat ttc ttc      450
Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala Leu Pro His Phe Phe
            105                 110                 115

atg gga tat tac agg tat tct aaa gaa act aat atc rat tca tca gaa      498
Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn Ile Xaa Ser Ser Glu
        120                 125                 130

aat tca aca tcg acc tta tcc act tgt tta att aat caa att tta tca      546
Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile Asn Gln Ile Leu Ser
    135                 140                 145

ctc aat aga gca tca cct gag ata gtg gga aaa ggt tgt tta aag gaa      594
Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys Gly Cys Leu Lys Glu
150                 155                 160                 165

tct ggg tca tac atg tgg ata tat gtg ttc atg ggt aat atg ctt cgt      642
Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met Gly Asn Met Leu Arg
                170                 175                 180

gga ata ggg gag act ccc ata gta cca ytg ggg ctt tct tac att gat      690
Gly Ile Gly Glu Thr Pro Ile Val Pro Xaa Gly Leu Ser Tyr Ile Asp
            185                 190                 195

gat ttc gct aaa gaa gga cat tct tct ttg tat tta ggt ata ttg aat      738
Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr Leu Gly Ile Leu Asn
        200                 205                 210

gca ata gca atg att ggt cca atc att ggc ttt acc ctg gga tct ctg      786
Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe Thr Leu Gly Ser Leu
    215                 220                 225

ttt tct aaa atg tac gtg gat att gga tat gta gat cta agc act atc      834
Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val Asp Leu Ser Thr Ile
230                 235                 240                 245

agg ata act cct act gat tct cga tgg gtt gga gct tgg tgg ctt aat      882
Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly Ala Trp Trp Leu Asn
                250                 255                 260
```

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ttc ctt gtg tct gga cta ttc tcc att att tct tcc ata cca ttc ttt      930
Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser Ser Ile Pro Phe Phe
            265                 270                 275

ttc ttg ccc caa act cca aat aaa cca caa aaa gaa aga aaa gct tca      978
Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys Glu Arg Lys Ala Ser
        280                 285                 290

ctg tct ttg cat gtg ctg gaa aca aat gat gaa aag gat caa aca gct     1026
Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu Lys Asp Gln Thr Ala
    295                 300                 305

aat ttg acc aat caa gga aaa aat att acc aaa aat gtg act ggt ttt     1074
Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys Asn Val Thr Gly Phe
310                 315                 320                 325

ttc cag tct ttt aaa agc atc ctt act aat ccc ctg tat gtt atg ttt     1122
Phe Gln Ser Phe Lys Ser Ile Leu Thr Asn Pro Leu Tyr Val Met Phe
                330                 335                 340

gtg ctt ttg acg ttg tta caa gta agc agc tat att ggt gct ttt act     1170
Val Leu Leu Thr Leu Leu Gln Val Ser Ser Tyr Ile Gly Ala Phe Thr
            345                 350                 355

tat gtc ttc aaa tac gta gag caa cag tat ggt cag cct tca tct aag     1218
Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly Gln Pro Ser Ser Lys
        360                 365                 370

gct aac atc tta ttg gga gtc ata acc ata cct att ttt gca agt gga     1266
Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro Ile Phe Ala Ser Gly
    375                 380                 385

atg ttt tta gga gga tat atc att aaa aaa ttc aaa ctg aac acc gtt     1314
Met Phe Leu Gly Gly Tyr Ile Ile Lys Lys Phe Lys Leu Asn Thr Val
390                 395                 400                 405

gga att gcc aaa ttc tca tgt ttt act gct gtg atg tca ttg tcc ttt     1362
Gly Ile Ala Lys Phe Ser Cys Phe Thr Ala Val Met Ser Leu Ser Phe
                410                 415                 420

tac cta tta tat ttt ttc ata ctc tgt gaa aac aaa tca gtt gcc gga     1410
Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn Lys Ser Val Ala Gly
            425                 430                 435

cta acc atg acc tat gat gga aat aat cca gtg aca tct cat aga gat     1458
Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val Thr Ser His Arg Asp
        440                 445                 450

gta cca ctt tct tat tgc aac tca gac tgc aat tgt gat gaa agt caa     1506
Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn Cys Asp Glu Ser Gln
    455                 460                 465

tgg gaa cca gtc tgt gga aac aat gga ata act tac atc tca ccc tgt     1554
Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr Tyr Ile Ser Pro Cys
470                 475                 480                 485

cta gca ggt tgc aaa tct tca agt ggc aat aaa aag cct ata gtg ttt     1602
Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys Lys Pro Ile Val Phe
                490                 495                 500
```

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tac aac tgc agt tgt ttg gaa gta act ggt ctc cag aac aga aat tac      1650
Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu Gln Asn Arg Asn Tyr
            505                 510                 515

tca gcc cat ttg ggt gaa tgc cca aga gat gat gct tgt aca agg aaa      1698
Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp Ala Cys Thr Arg Lys
        520                 525                 530

ttt tac ttt ttt gtt gca ata caa gtc ttg aat tta ttt ttc tct gca      1746
Phe Tyr Phe Phe Val Ala Ile Gln Val Leu Asn Leu Phe Phe Ser Ala
    535                 540                 545

ctt gga ggc acc tca cat gtc atg ctg att gtt aaa att gtt caa cct      1794
Leu Gly Gly Thr Ser His Val Met Leu Ile Val Lys Ile Val Gln Pro
550                 555                 560                 565

gaa ttg aaa tca ctt gca ctg ggt ttc cac tca atg gtt ata cga gca      1842
Glu Leu Lys Ser Leu Ala Leu Gly Phe His Ser Met Val Ile Arg Ala
                570                 575                 580

cta gga gga att cta gct cca ata tat ttt ggg gct ctg att gat aca      1890
Leu Gly Gly Ile Leu Ala Pro Ile Tyr Phe Gly Ala Leu Ile Asp Thr
            585                 590                 595

acg tgt ata aag tgg tcc acc aac aac tgt ggc aca cgt ggg tca tgt      1938
Thr Cys Ile Lys Trp Ser Thr Asn Asn Cys Gly Thr Arg Gly Ser Cys
        600                 605                 610

agg aca tat aat tcc aca tca ttt tca agg gtc tac ttg ggc ttg tct      1986
Arg Thr Tyr Asn Ser Thr Ser Phe Ser Arg Val Tyr Leu Gly Leu Ser
    615                 620                 625

tca atg tta aga gtc tca tca ctt gtt tta tat att ata tta att tat      2034
Ser Met Leu Arg Val Ser Ser Leu Val Leu Tyr Ile Ile Leu Ile Tyr
630                 635                 640                 645

gcc atg aag aaa aaa tat caa gag aaa gat atc aat gca tca gaa aat      2082
Ala Met Lys Lys Lys Tyr Gln Glu Lys Asp Ile Asn Ala Ser Glu Asn
                650                 655                 660

gga agt gtc atg gat gaa gca aac tta gaa tcc tta aat aaa aat aaa      2130
Gly Ser Val Met Asp Glu Ala Asn Leu Glu Ser Leu Asn Lys Asn Lys
            665                 670                 675

cat ttt gtc cct tct gct ggg gca gat agt gaa aca cat tgt taa          2175
His Phe Val Pro Ser Ala Gly Ala Asp Ser Glu Thr His Cys  *
        680                 685                 690

ggggagaaaa aaagccactt ctgcttctgt gtttccaaac agcattgcat tgattcagta   2235
agatgttatt tttgaggagt tcctggtcct ttcactaaga atttccacat cttttatggt   2295
ggaagtataa ataagcctat gaacttataa taaaacaaac tgtaggtaga aaaaatgaga   2355
gtactcattg ttacattata gctacatatt tgtggttaag gttagactat atgatccata   2415
caaattaaag tgagagacat ggttactgtg taataaaaga aaaaatactt gttcaggtaa   2475
ttctaattct taataaaaca aatgagtatc atacaggtag aggttaaaaa ggaggagcta   2535
gattcatatc ctaagtaaag agaaatgcct agtgtctatt ttattaaaca aacaaacaca   2595
gagtttgaac tataatacta aggcctgaag tctagcttgg atatatgcta caataatatc   2655
tgttactcac ataaaattat atatttcaca gactttatca atgtataatt aacaattatc   2715
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tttgcaaaaa aaaaaaaaaa aaaaa                                         2800
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<210> 6
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<213> H. sapiens

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<222> (130)...(130)
<223> Xaa = Asp or Asn

<221> VARIANT
<222> (191)...(191)
<223> Xaa = Leu

<400> 6

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Glu Asn Lys Lys Thr Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala
            20                  25                  30
Ala Leu Ser Leu Ser Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met
        35                  40                  45
Lys Ser Ser Ile Ile His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser
    50                  55                  60
Leu Val Gly Phe Ile Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val
65                  70                  75                  80
Ile Val Phe Val Ser Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu
                85                  90                  95
Ile Gly Ile Gly Cys Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala
            100                 105                 110
Leu Pro His Phe Phe Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn
        115                 120                 125
Ile Xaa Ser Ser Glu Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile
    130                 135                 140
Asn Gln Ile Leu Ser Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys
145                 150                 155                 160
Gly Cys Leu Lys Glu Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met
                165                 170                 175
Gly Asn Met Leu Arg Gly Ile Gly Glu Thr Pro Ile Val Pro Xaa Gly
            180                 185                 190
Leu Ser Tyr Ile Asp Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr
        195                 200                 205
Leu Gly Ile Leu Asn Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe
    210                 215                 220
Thr Leu Gly Ser Leu Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val
225                 230                 235                 240
Asp Leu Ser Thr Ile Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly
                245                 250                 255
Ala Trp Trp Leu Asn Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser
            260                 265                 270
Ser Ile Pro Phe Phe Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys
        275                 280                 285
Glu Arg Lys Ala Ser Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu
    290                 295                 300
Lys Asp Gln Thr Ala Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys
305                 310                 315                 320
Asn Val Thr Gly Phe Phe Gln Ser Phe Lys Ser Ile Leu Thr Asn Pro
                325                 330                 335
Leu Tyr Val Met Phe Val Leu Leu Thr Leu Leu Gln Val Ser Ser Tyr
            340                 345                 350
Ile Gly Ala Phe Thr Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly
        355                 360                 365
Gln Pro Ser Ser Lys Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro
    370                 375                 380
```

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Ile Phe Ala Ser Gly Met Phe Leu Gly Gly Tyr Ile Ile Lys Lys Phe
385                 390                 395                 400
Lys Leu Asn Thr Val Gly Ile Ala Lys Phe Ser Cys Phe Thr Ala Val
                405                 410                 415
Met Ser Leu Ser Phe Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn
            420                 425                 430
Lys Ser Val Ala Gly Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val
        435                 440                 445
Thr Ser His Arg Asp Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn
    450                 455                 460
Cys Asp Glu Ser Gln Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr
465                 470                 475                 480
Tyr Ile Ser Pro Cys Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys
                485                 490                 495
Lys Pro Ile Val Phe Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu
            500                 505                 510
Gln Asn Arg Asn Tyr Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp
        515                 520                 525
Ala Cys Thr Arg Lys Phe Tyr Phe Phe Val Ala Ile Gln Val Leu Asn
    530                 535                 540
Leu Phe Phe Ser Ala Leu Gly Gly Thr Ser His Val Met Leu Ile Val
545                 550                 555                 560
Lys Ile Val Gln Pro Glu Leu Lys Ser Leu Ala Leu Gly Phe His Ser
                565                 570                 575
Met Val Ile Arg Ala Leu Gly Gly Ile Leu Ala Pro Ile Tyr Phe Gly
            580                 585                 590
Ala Leu Ile Asp Thr Thr Cys Ile Lys Trp Ser Thr Asn Asn Cys Gly
        595                 600                 605
Thr Arg Gly Ser Cys Arg Thr Tyr Asn Ser Thr Ser Phe Ser Arg Val
    610                 615                 620
Tyr Leu Gly Leu Ser Ser Met Leu Arg Val Ser Ser Leu Val Leu Tyr
625                 630                 635                 640
Ile Ile Leu Ile Tyr Ala Met Lys Lys Lys Tyr Gln Glu Lys Asp Ile
                645                 650                 655
Asn Ala Ser Glu Asn Gly Ser Val Met Asp Glu Ala Asn Leu Glu Ser
            660                 665                 670
Leu Asn Lys Asn Lys His Phe Val Pro Ser Ala Gly Ala Asp Ser Glu
        675                 680                 685
Thr His Cys
    690
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<210> 7
<211> 2360
<212> DNA
<213> H. sapiens

<220>
<221> CDS
<222> (100)...(1729)
<223> Coding sequence ATnov3.2

<221> variation
<222> (1705)...(1710)
<223> Polymorphism of 5 or 6 T residues

<221> variation
<222> (487)...(487)
<223> Polymorphism of A or G

<221> variation
<222> (670)...(670)
<223> Polymorphism of C or T

<400> 7

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gtggacttgt tgcagttgct gtaggattct aaatccaggt gattgtttca aactgagcat      60
caacaacaaa aacatttgta tgatatctat atttcaatc atg gac caa aat caa         114
                                        Met Asp Gln Asn Gln
                                         1               5

cat ttg aat aaa aca gca gag gca caa cct tca gag aat aag aaa aca         162
His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser Glu Asn Lys Lys Thr
                10                  15                  20

aga tac tgc aat gga ttg aag atg ttc ttg gca gct ctg tca ctc agc         210
Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala Ala Leu Ser Leu Ser
            25                  30                  35

ttt att gct aag aca cta ggt gca att att atg aaa agt tcc atc att         258
Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met Lys Ser Ser Ile Ile
        40                  45                  50

cat ata gaa cgg aga ttt gag ata tcc tct tct ctt gtt ggt ttt att         306
His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser Leu Val Gly Phe Ile
    55                  60                  65

gac gga agc ttt gaa att gga aat ttg ctt gtg att gta ttt gtg agt         354
Asp Gly Ser Phe Glu Ile Gly Asn Leu Leu Val Ile Val Phe Val Ser
70                  75                  80                  85

tac ttt gga tcc aaa cta cat aga cca aag tta att gga atc ggt tgt         402
Tyr Phe Gly Ser Lys Leu His Arg Pro Lys Leu Ile Gly Ile Gly Cys
                90                  95                  100

ttc att atg gga att gga ggt gtt ttg act gct ttg cca cat ttc ttc         450
Phe Ile Met Gly Ile Gly Gly Val Leu Thr Ala Leu Pro His Phe Phe
            105                 110                 115

atg gga tat tac agg tat tct aaa gaa act aat atc rat tca tca gaa         498
Met Gly Tyr Tyr Arg Tyr Ser Lys Glu Thr Asn Ile Xaa Ser Ser Glu
        120                 125                 130

aat tca aca tcg acc tta tcc act tgt tta att aat caa att tta tca         546
Asn Ser Thr Ser Thr Leu Ser Thr Cys Leu Ile Asn Gln Ile Leu Ser
    135                 140                 145

ctc aat aga gca tca cct gag ata gtg gga aaa ggt tgt tta aag gaa         594
Leu Asn Arg Ala Ser Pro Glu Ile Val Gly Lys Gly Cys Leu Lys Glu
150                 155                 160                 165

tct ggg tca tac atg tgg ata tat gtg ttc atg ggt aat atg ctt cgt         642
Ser Gly Ser Tyr Met Trp Ile Tyr Val Phe Met Gly Asn Met Leu Arg
                170                 175                 180

gga ata ggg gag act ccc ata gta cca ytg ggg ctt tct tac att gat         690
Gly Ile Gly Glu Thr Pro Ile Val Pro Xaa Gly Leu Ser Tyr Ile Asp
            185                 190                 195

gat ttc gct aaa gaa gga cat tct tct ttg tat tta ggt ata ttg aat         738
Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr Leu Gly Ile Leu Asn
        200                 205                 210

gca ata gca atg att ggt cca atc att ggc ttt acc ctg gga tct ctg         786
Ala Ile Ala Met Ile Gly Pro Ile Ile Gly Phe Thr Leu Gly Ser Leu
    215                 220                 225
```

```
ttt tct aaa atg tac gtg gat att gga tat gta gat cta agc act atc      834
Phe Ser Lys Met Tyr Val Asp Ile Gly Tyr Val Asp Leu Ser Thr Ile
230                 235                 240                 245

agg ata act cct act gat tct cga tgg gtt gga gct tgg tgg ctt aat      882
Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly Ala Trp Trp Leu Asn
                250                 255                 260

ttc ctt gtg tct gga cta ttc tcc att att tct tcc ata cca ttc ttt      930
Phe Leu Val Ser Gly Leu Phe Ser Ile Ile Ser Ser Ile Pro Phe Phe
            265                 270                 275

ttc ttg ccc caa act cca aat aaa cca caa aaa gaa aga aaa gct tca      978
Phe Leu Pro Gln Thr Pro Asn Lys Pro Gln Lys Glu Arg Lys Ala Ser
        280                 285                 290

ctg tct ttg cat gtg ctg gaa aca aat gat gaa aag gat caa aca gct     1026
Leu Ser Leu His Val Leu Glu Thr Asn Asp Glu Lys Asp Gln Thr Ala
    295                 300                 305

aat ttg acc aat caa gga aaa aat att acc aaa aat gtg act ggt ttt     1074
Asn Leu Thr Asn Gln Gly Lys Asn Ile Thr Lys Asn Val Thr Gly Phe
310                 315                 320                 325

ttc cag tct ttt aaa agc atc ctt act aat ccc ctg tat gtt atg ttt     1122
Phe Gln Ser Phe Lys Ser Ile Leu Thr Asn Pro Leu Tyr Val Met Phe
                330                 335                 340

gtg ctt ttg acg ttg tta caa gta agc agc tat att ggt gct ttt act     1170
Val Leu Leu Thr Leu Leu Gln Val Ser Ser Tyr Ile Gly Ala Phe Thr
            345                 350                 355

tat gtc ttc aaa tac gta gag caa cag tat ggt cag cct tca tct aag     1218
Tyr Val Phe Lys Tyr Val Glu Gln Gln Tyr Gly Gln Pro Ser Ser Lys
        360                 365                 370

gct aac atc tta ttg gga gtc ata acc ata cct att ttt gca agt gga     1266
Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro Ile Phe Ala Ser Gly
    375                 380                 385

atg ttt tta gga gga tat atc att aaa aaa ttc aaa ctg aac acc gtt     1314
Met Phe Leu Gly Gly Tyr Ile Ile Lys Lys Phe Lys Leu Asn Thr Val
390                 395                 400                 405

gga att gcc aaa ttc tca tgt ttt act gct gtg atg tca ttg tcc ttt     1362
Gly Ile Ala Lys Phe Ser Cys Phe Thr Ala Val Met Ser Leu Ser Phe
                410                 415                 420

tac cta tta tat ttt ttc ata ctc tgt gaa aac aaa tca gtt gcc gga     1410
Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn Lys Ser Val Ala Gly
            425                 430                 435

cta acc atg acc tat gat gga aat aat cca gtg aca tct cat aga gat     1458
Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val Thr Ser His Arg Asp
        440                 445                 450

gta cca ctt tct tat tgc aac tca gac tgc aat tgt gat gaa agt caa     1506
Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn Cys Asp Glu Ser Gln
    455                 460                 465
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tgg gaa cca gtc tgt gga aac aat gga ata act tac atc tca ccc tgt      1554
Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr Tyr Ile Ser Pro Cys
470                 475                 480                 485

cta gca ggt tgc aaa tct tca agt ggc aat aaa aag cct ata gtg ttt      1602
Leu Ala Gly Cys Lys Ser Ser Ser Gly Asn Lys Lys Pro Ile Val Phe
                490                 495                 500

tac aac tgc agt tgt ttg gaa gta act ggt ctc cag aac aga aat tac      1650
Tyr Asn Cys Ser Cys Leu Glu Val Thr Gly Leu Gln Asn Arg Asn Tyr
            505                 510                 515

tca gcc cat ttg ggt gaa tgc cca aga gat gat gct tgt aca agg aaa      1698
Ser Ala His Leu Gly Glu Cys Pro Arg Asp Asp Ala Cys Thr Arg Lys
        520                 525                 530

ttt tac ttt ttg ttg caa tac aag tct tga a tttatttttc tctgcacttg      1749
Phe Tyr Phe Leu Leu Gln Tyr Lys Ser  *
    535                 540

gaggcacctc acatgtcatg ctgattgtta aaattgttca acctgaattg aaatcacttg    1809
cactgggttt ccactcaatg gttatacgag cactaggagg aattctagct ccaatatatt    1869
ttggggctct gattgataca acgtgtataa agtggtccac caacaactgt ggcacacgtg    1929
ggtcatgtag gacatataat tccacatcat tttcaagggt ctacttgggc ttgtcttcaa    1989
tgttaagagt ctcatcactt gttttatata ttatattaat ttatgccatg aagaaaaaat    2049
atcaagagaa agatatcaat gcatcagaaa atggaagtgt catggatgaa gcaaacttag    2109
aatccttaaa taaaaataaa cattttgtcc cttctgctgg ggcagatagt gaaacacatt    2169
gttaagggga gaaaaaaagc cacttctgct tctgtgtttc caaacagcat tgcattgatt    2229
cagtaagatg ttatttttga ggagttcctg gtcctttcac taagaatttc cacatctttt    2289
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aaaaaaaaaa a                                                        2360
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<210> 8
<211> 542
<212> PRT
<213> H. sapiens

<220>
<221> VARIANT
<222> (130)...(130)
<223> Xaa = Asp or Asn

<221> VARIANT
<222> (191)...(191)
<223> Xaa = Leu

<400> 8

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Met Asp Gln Asn Gln His Leu Asn Lys Thr Ala Glu Ala Gln Pro Ser
 1               5                  10                  15
Glu Asn Lys Lys Thr Arg Tyr Cys Asn Gly Leu Lys Met Phe Leu Ala
            20                  25                  30
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ttt att gct aag aca cta ggt gca att att atg aaa agt tcc atc att         258
Phe Ile Ala Lys Thr Leu Gly Ala Ile Ile Met Lys Ser Ser Ile Ile
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cat ata gaa cgg aga ttt gag ata tcc tct tct ctt gtt ggt ttt att         306
His Ile Glu Arg Arg Phe Glu Ile Ser Ser Ser Leu Val Gly Phe Ile
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Asp Phe Ala Lys Glu Gly His Ser Ser Leu Tyr Leu Gly Ile Leu Asn
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230                 235                 240                 245

agg ata act cct act gat tct cga tgg gtt gga gct tgg tgg ctt aat      882
Arg Ile Thr Pro Thr Asp Ser Arg Trp Val Gly Ala Trp Trp Leu Asn
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gct aac atc tta ttg gga gtc ata acc ata cct att ttt gca agt gga     1266
Ala Asn Ile Leu Leu Gly Val Ile Thr Ile Pro Ile Phe Ala Ser Gly
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Tyr Leu Leu Tyr Phe Phe Ile Leu Cys Glu Asn Lys Ser Val Ala Gly
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cta acc atg acc tat gat gga aat aat cca gtg aca tct cat aga gat      1458
Leu Thr Met Thr Tyr Asp Gly Asn Asn Pro Val Thr Ser His Arg Asp
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gta cca ctt tct tat tgc aac tca gac tgc aat tgt gat gaa agt caa      1506
Val Pro Leu Ser Tyr Cys Asn Ser Asp Cys Asn Cys Asp Glu Ser Gln
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Trp Glu Pro Val Cys Gly Asn Asn Gly Ile Thr Tyr Ile Ser Pro Cys
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tca gcc cat ttg ggt gaa tgc cca aga gat gat gct tgt aca agg aaa      1698
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acg tgt ata aag tgg tcc acc aac aac tgt ggc aca cgt ggg tca tgt      1938
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(57) Abstract

Methods for isolating ATnov genes are provided. The ATnov nucleic acid compositions find use in identifying homologous or related proteins and the DNA sequences encoding such proteins; in producing compositions that modulate the expression or function of the protein; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as identification of cell type based on expression, and the like.

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UA	Ukraine
UG	Uganda
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UZ	Uzbekistan
VN	Viet Nam
YU	Yugoslavia
ZW	Zimbabwe

INTERNATIONAL SEARCH REPORT

International Application No
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A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/12 C12N5/10 C12Q1/68 A01K67/00 C12N15/00
C07K16/28 C07K14/705

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 C12N C07K A61K C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	KULLAK-UBLICK ET AL: "Molecular and functional characterization of an organic anion transporting polypeptide cloned from human liver" GASTROENTEROLOGY, vol. 109, 1 October 1995 (1995-10-01), pages 1274-1282, XP002092892 ISSN: 0016-5085 cited in the application the whole document ---	1
A	WO 97 31111 A (INTROGENE BV ;AMC AMSTERDAM (NL); KANKER INST NL (NL); OUDE ELFERI) 28 August 1997 (1997-08-28) the whole document ---	1
	-/--	

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
9 March 2000	13.07.00
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer CHAMBONNET, F

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Form PCT/ISA/210 (second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

International Application No
PC./US 99/17823

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	Embase accession number AB026257.1 Created 08-JUN-1999 , Sequence characterization: Homo sapiens mRNA for organic anion transporter OATP-C, complete XP002132655 the whole document ---	1-3
T	HSIANG B.H., ZHU Y., WANG Z., WU Y., SASSEVILLE V., YANG W.-P.,: "A novel human hepatic organic anion transporting polypeptide (OATP2) Identification of a liver-specific human organic anion transporting polypeptide and identification of rat and human hydroxymethylglutaryl-coa reductase inhibitor transporter." JOURNAL OF BIOLOGICAL CHEMISTRY (MICROFILMS), vol. 274, no. 52, 24 December 1999 (1999-12-24), pages 37161-37168, XP002131865 MD US the whole document ---	1-3
T	ABE T., ET AL.: "Identification of a Novel Gene Family Encoding Human Liver-specific Organic Anion Transporter LST-1" J. BIOL. CHEM., vol. 274, no. 24, 11 June 1999 (1999-06-11), pages 17159-17163, XP000877288 the whole document -----	1-4

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 99/17823

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Claims 2, 3, 15-17 completely, and 1, 4-14 partially.

Remark on Protest

☐ The additional search fees were accompanied by the applicant's protest.

☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

1. Claims: 2 3 15-17 and partially 1 4-14

An isolated nucleic acid comprising a nucleotide sequence selected from the group of SEQ NO:3, SEQ NO:5, SEQ NO:7, and SEQ NO:9, or encoding a mammalian ATnov3 protein, having an amino acid sequence selected from the group of SEQ NO:4, SEQ NO:6, SEQ NO:8 and SEQ NO:10; an expression cassette comprising it; a cell comprising said expression cassette; a method for producing a protein using said cell; mammalian ATnov3 polypeptide; a monoclonal antibody binding to said protein; a non human transgenic animal model for ATnov3 gene function; an isolated nucleic acid probe comprising an ATnov3 sequence polymorphism; an array of oligonucleotides comprising probes for detection of ATnov3 locus polymorphisms.

2. Claims: partially 1 4-14

An isolated nucleic acid comprising the nucleotide sequence as set forth in SEQ NO:1 or encoding a mammalian ATnov1 protein; an expression cassette comprising it; a cell comprising said expression cassette; a method for producing a protein using said cell; mammalian ATnov1 polypeptide; a monoclonal antibody binding to said protein; a non human transgenic animal model for ATnov1 gene function;

3. Claims: partially 1 4-14

An isolated nucleic acid comprising the nucleotide sequence as set forth in SEQ NO:2 or encoding a mammalian ATnov2 protein; an expression cassette comprising it; a cell comprising said expression cassette; a method for producing a protein using said cell; mammalian ATnov2 polypeptide; a monoclonal antibody binding to said protein; a non human transgenic animal model for ATnov2 gene function;

INTERNATIONAL SEARCH REPORT

formation on patent family members

International Application No
Pt /US 99/17823

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9731111 A	28-08-1997	AU 1736697 A	10-09-1997
		EP 0900273 A	10-03-1999